PHYTOCHEMICAL INVESTIGATION AND IN VITRO CONSERVATION OF AMARANTHUS VIRIDIS L. ALEAFY VEGETABLE OF KERALA.

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Abstract

Antioxidant compounds in food play an important role as a health protecting factor. Primary sources of naturally occurring antioxidants are whole grains, fruits and vegetables. Highly reactive free radicals and oxygen species can initiate degenerative diseases. Amaranthus viridis of Amaranthaceae is a wild species used by tribal people of Kerala for treating various diseases and for culinary purposes in other countries. The plant exhibits sufficient characteristics related to claims of its therapeutic properties. Crude methanolic leaf extract is used for preliminary phytochemical analysis. Phytochemicals detected in the study can have direct or indirect effects on properties attributed to the plant. Quantitative phytochemical analysis showed higher amounts of total carbohydrate, total proteins in addition to a considerable amount of reducing sugar, starch and pigments like chlorophyll-a, chlorophyll-b and Total chlorophyll. Antioxidant activity of the leaf was evaluated for different enzymatic and non- enzymatic antioxidants. Total polyphenol and superoxide dismutase were present in higher amounts, satisfying its use as a potential source of antioxidants. Anticancer analysis in EAC (Ehrlich's Ascites carcinoma) and DLA (Dalton's Lymphoma Ascites) showed Higher cytotoxicity in EAC compared to DLA against standard revealing promising anticancer effects of the leaf extract. In vitro conservation of different explants on MS medium supplemented 2 mg/L BAP showed bare survival capacity of nodal explant with axillary bud. Present study evaluated various nutritional, medicinal and regenerative aspects of the plant Amaranthus viridis, exhibiting diverse potentialities of the plant and providing supporting information for its use as an ethnomedicinal plant.

Keywords: Amaranthus viridis, Amaranthaceae, Ethnomedicinal plant, EAC, DLA and in vitro conservation

Introduction

Leafy vegetables played crucial roles in complementing diets for humans and animals (Amadiet al., 2013). Amaranthus viridis is sometimes eaten as cooked vegetable, fodder for cattle and green manure; the leaves are diuretic and purgative and are used for treating inflammations, boils and abscesses, gonorrhoea, orchitis and haemorrhoids. (Alegbejo, 2013). Amaranthus viridis L., the main type of vegetable amaranth, seems to have originated in south or south-east Materials and Methods Asia, and is consequently dispersed throughout Collection and Preparation of Sample for the tropical and temperate regions (Martin and Phytochemical Analysis. Telek, 1979). Boiled leaves and roots of Ama- The plant Amaranthus viridis was collected as ranthus are used as laxative, diuretic, anti- fresh from Kalady, Thiruvananthapuram district diabetic, antipyretic, anti-snake venom, to re- and Panmana, Kollam district of Kerala. The lieve breathing in acute bronchitis. It also has plant specimen was made into herbarium and anti-inflammatory properties, immunomodula- deposited in the herbarium repository of Botanitory activity, anti-androgenic activity and cal Survey of India (BSI), Southern Regional

anthelmintic properties (Janet, 2013). To address this lacuna, the present study was carried out for preliminary phytochemical analysis, evaluation of nutritional properties, evaluation of antioxidant properties and evaluate anticancer property of the crude methanol extract in DLA and EAC cell line and in vitro conservation of plant. In future Amaranthusviridis is important natural source of developing new drugs. So have select the present plant for this study.

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Centre (SRC), Coimbatore-3 and authenticated The fresh leaves of Amaranthus viridis were BSI/SRC/5/23/2023-24/Tech-466. No.: sample preparation, fresh leaves and stem were and experiment was repeated thrice to confirm separated, shade dried, ground well using a me- the result. The analysis was preformed followchanical blender to fine powder and transferred ing standard methods for estimation of reducing into airtight containers for further analysis.

Phytochemical Screening Preparation of Plant Extract

methanol for 8 hours by Soxhlet apparatus and pene (Zakaria et al., 1979), carotenoids (Bendich extracted as green, black solid respectively. Af- and Olson, 1989), total polyphenol (Eom et al., ter which, the residues were transferred to a pre- 2008), polyphenol oxidase (Esterbauer et weighted sample container for storage and later al., 1991) superoxide dismutase (Gong et al., used for phytochemical screening. The phyto- 2005) and Lipid peroxide (Ewa, 2006). chemicals like reducing sugar, flavonoids, alkaloids, tannins, terpenoids, steroids, saponins and Results and Discussion anthraquinones were tested (Pradeesh and Fresh leaves and stems of Amaranthus viridis Swapna, 2018).

Preparation of Plant Extract

methanol for 8 hours by Soxhlet apparatus and explants of the plant were used to carry out in extracted as green, black solid respectively. Af- vitro conservation. ter which, the residues were transferred to a preweighted sample container for storage and later Phytochemicals screened in the present study used for phytochemical screening.

Phytochemical Screening

Phytochemical analysis of plant extract was done as described by Harborne (Harborne, 1977). The different phytochemicals like reducing sugar, flavonoids, alkaloids, tannins, terpenoids, steroids, saponins and anthraquinones were tested.

Biochemical analysis

For used for the nutritional and antioxidant analysis sugar (Miller, 1972), total carbohydrate (Hedge and Hofreiter, 1962), total protein (Lowry et al., 1951), chlorophyll (Witham et al., 1971), starch (Thayumanavan and Sadasivam, 1984) and anti-The dried plant materials were extracted with oxidant like proline (Bates et al., 1973), lyco-

were studied for its nutritional and antioxidant properties. Crude methanolic extract was used for the preliminary phytochemical investigation The dried plant materials were extracted with and in vitro anticancer analysis. The different

> include reducing sugar, flavonoids, alkaloids, tannins, terpenoids, steroids, anthraquinones. The concentrations of tannins and saponins were high for both leaves and stem of Amaranthus viridis. While the concentrations of reducing sugars, flavonoids, alkaloids, steroids, anthraquinones are very low concentration, terpenoids were not found in Amaranthus viridis leaf extract. The concentrations of steroids, terpenoids and anthraquinones were not found in Amaranthus viridis stem extract (Table 1).

Sl No:	Phytochemicals	A.viridis Leaf	A.viridis Stem
1	Reducing sugar	+	+
2	Flavonoids	+	+
3	Alkaloids	+	+
4	Tannins	++	++
5	Terpenoids	-	-
6	Steroids	-	
7	Saponins	++	++
8	Anthraquinones	+	-

Table 1. Preliminary Phytochemical Evaluation of Amaranthus viridis

Ouantitative Analysis Nutritional Evaluation

Plant extract has a significant effect in reducing hydrates, proteins, pigments and starch. blood sugar level and therefore can be used as an alternative and effective insulin enhancer for Evaluation of Antioxidant Properties individuals who have a symptom of diabetes Plants are the reservoir of antioxidants that can mellitus (Floramyr, 2019). Reducing sugar from act as an endogenous antioxidant source through the aerial plant parts were extracted and ana- diet which can help in resolving free radical melysed by Dinitro salicylic acid method and the diated health conditions and its degenerative efresults were found 0.982 mg g⁻¹ in leaves of fects in human beings. Evaluation of enzymatic Amaranthus viridis and 0.912 mg g⁻¹ in the stem and non-enzymatic antioxidants in Amaranthus of Amaranthus viridis (Fig.1). Carbohydrates viridis can help in understanding the therapeutic are macronutrients present in plants that forms potential of the plant in terms of its antioxidant major constituents of human diet that can be me- properties. Present study evaluated tabolized to yield energy (Pradeesh and enzymatic antioxidants like proline, lycopene, Praveena, 2020). Present study estimated total carotenoids and polyphenols and enzymatic ancarbohydrates present in leaf tissue using the tioxidants such as superoxide dismutase (SOD), anthrone method. The amount of carbohydrates polyphenol oxidase (PPO), amylase and lipid in leaves of Amaranthus viridis was found to be peroxidase (LPx) by standard estimation methhigher 35.908 mgg⁻¹ and in stem 33.128 mgg⁻¹ ods. Proline is an amino acid molecule that as shown in (figure 2).

Proteins are complex macromolecules that have death and gene expression) in plants (Szabados structural, functional and catalytic roles in or- and Arnould, 2010). The amount of proline in ganisms with amino acids as structural units. Amaranthus viridis was estimated 0.618 mg g They form the building block of tissue. (Ghali ¹in leaf and 0.519 mg g⁻¹ in stem (Fig. 5). Lycoand Alkoaik, 2010). Estimation of protein from pene comprises the most important class of cathe leaves of Amaranthus viridis was done by rotenoids which at physiological concentration Lowry's method and the result was found to be has the potential to act against cancerous cells high in leaves (32.719 mg g⁻¹) and stem (30.921 and its proliferation. It interferes with growth mg g⁻¹) as shown in Fig. 2. Dietary chlorophyll factor receptor signalling and cell cycle progresderivative has the ability to scavenge long lived sion in prostate cancer cells without toxic effects free radicals such as DPPH and ABTS (Mishra and cell apoptosis (David and Lu, 2002). The et al., 2011). The estimation of Chlorophyll in estimated amount of proline in methanolic leaf Amaranthus viridis was carried out using Ar- extract of Amaranthus viridis (0.152 mg g⁻¹) and non's method with a result of high content of in stem extract of Amaranthus viridis (0.082 mg chlorophyll-a (0.898 mg g^{-1}), chlorophyll-b g^{-1}) as shown in figure 5. $(0.862 \text{ mg g}^{-1})$ and total chlorophyll (1.436 mg g) $^{-1}$) in leaves and chlorophyll-a (0.628 mg g⁻¹), Carotenoids are widely distributed lipophilic chlorophyll-b (0.592 mg g⁻¹) and total chloro- pigment molecules with antioxidant properties. phyll (1.136 mg g⁻¹) in stem (Fig. 3). Commer- They act as antenna molecules in capturing light cial production of starch can be done by degra- and in transfer of energy to chlorophyll moledation of starch. It is used as thickening agents cules during photosynthesis. They also give proin sauces and as colloidal stabilisers in salad tection to the photosystem by interacting with dressings (Manthey and Xu, 2009). The amount free radicals and to light induced tissue damage. of starch in Amaranthus viridis was found to be Carotenoids can quench highly reactive singlet 0.832 mg g⁻¹ in leaves and 0.631 mg g⁻¹ in stem oxygen and block free radical mediated reacas shown in Fig. 4. The nutritional analysis of tions (Bendich and Olson, 1989). The estimated

leaves of Amaranthus viridis showed the presence of a high amount of reducing sugar, carbo-

nonplays an important role in physiological as well as cellular processes (cell proliferation, cell

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amount of carotenoids in leaves of Amaranthus currence of chain reactions changes lipid struc*viridis* is 0.418 mg g^{-1} and in stem 0.396 mg g^{-1} ture and organisation of cell membrane in the (Fig.5). Flavonoids are a type of secondary me- process of lipid peroxidation. Peroxidase entabolites obtained from plants. It is usually zymes (stress enzymes) are hemi-containing enfound in fruits, vegetables, nuts, seeds, stems zymes that can oxidise various substrates using and flowers. Flavonoids are also the best sources H₂O₂ that prevent its accumulation under menatural antioxidants of (Sankhadipet al., 2018). The amount of total fla- venting lipid peroxidation (Pradeesh and vonoids is 0.920 mg g⁻¹ in the leaves and 0.892 Swapna, 2018). The result revealed that the mg g^{-1} in the stem of *Amaranthus viridis* and amount of enzymatic antioxidant lipoperoxide is found to be high (Fig .5).

Superoxide dismutase (SOD) are metalloenzymes catalysing dismutation or reaction of su- Evaluation of Pharmacological Property peroxide radicals. On the basis of the cofactor Plants have been known to be effective in treatpresent, they are classified into Fe-SOD (found ing various diseases since ancient times. Cancer in mitochondria) and Cu-SOD or Zn-SOD is one of the leading fatal diseases of man. The (found in chloroplast, peroxisomes and cytosol). relevance of active compounds presents in They are activated under different abiotic stress plants in treating malignant tumours and preconditions such as water deficiency, chilling, venting cancer is acquiring more popularity in heat, hypoxia and Heavy metals in toxic concen- recent years. Present study evaluated in vitro trations. Intracellular SOD may play a key role anticancer activity of Amaranthus viridis leaf in protection of cancer cells against reactive extract and stem extract in methanol. Anticancer oxygen species generated by anticancer drugs effect was analysed using Dalton's Lymphoma and radiation (Shingo et al., 1994). The amount Ascites (DLA) and Ehrlich Ascites Carcinoma of superoxide dismutase is 1.296 mg g⁻¹ leaves and 1.819 mg g^{-1} in the stem of *Amaran*- Trypan blue exclusion method. The viable cell *thus viridis* as shown in Fig .6. Polyphenol oxi- suspension 1×10^6 cells in 0.1 ml was added in dases (PPO) mediated reactions play an impor- the tubes containing various concentrations tant role in alteration of colour, flavour, texture (100, 500 and 1000 µg/ml) of test compounds and nutritional values of fruit and vegetable and the volume was made up to 1 ml using crops (Yurok and Marshall, 2003). The esti- phosphate buffer saline (PBS). The mixtures mated amount of enzymatic antioxidant poly- were incubated for 3 hours at 37°C and were phenol oxidase is 0.956 mg g^{-1} in leaf and 0.813 added with 2 drops of Trypan blue dye. Dead mg g⁻¹ in stem of *Amaranthus viridis* (Fig 6.). cells take up the blue colour of the dye while the Amylases are one of the most thoroughly re- live cells do not. Reduction in the viable cell searched digestive enzymes catalysing degrada- count and increased non-viable cancer cell count tion of polysaccharides or reserve carbohydrates towards normal in tumour host suggest antitu-(starch in plants) to simple sugars providing en- mor effect against EAC and DLA cells in mice. ergy source in organisms. Amylase play an im- Cyclophosphamide is used as a standard antiportant role in inducing growth of embryos by cancer compound. The result obtained from the breakdown of starch to sugar in the seeds anticancer study revealed that the leaf methanol (Pradeesh and Swapna, 2018). The estimated extract of Amaranthus viridisshowed 46.2, 73.9, amount of enzymatic antioxidant amylases is 86.4 cytotoxicity in EAC compared to 41.9, 0.948 mg g⁻¹ in leaf and 0.892 mg g⁻¹ in stem of 66.9, 78.2 cytotoxicity in DLA and the stem Amaranthus viridis as shown in Fig 6. Free radi- methanol extract showed 39.8, 60.3, 79.4 cytocals such as H₂O₂ attacks unsaturated fatty acids toxicity in EAC compared to 35.6, 49.2, 62.1 producing lipid hydro peroxides and further oc- cytotoxicity in DLA at concentrations of 100,

in human diets tabolism during stress conditions thereby pre-0.912 mg g^{-1} in leaves and 0.826 mg g^{-1} in the stem of Amaranthus viridis (Fig 6.).

in (EAC) cell lines. Viability was determined by

500 and 1000 µg/ml (Fig 7.) and (Table 2.). Re- tract can provide possibilities to novel therapeusults obtained in the present study demonstrated tic findings for treating cancer cells. that the methanol extract of the Amaranthus viridis exhibits in vitro anticancer activity against In vitro Conservation of Nodal Explants of DLA and EAC cell lines. The leaf extract Amaranthus viridis showed concentration dependent cytotoxicity In vitro conservation of Amaranthus viridis was which was found to be effective against solid carried out by using different explants such as tumours induced by DLA and ascites tumours stem, node with axillary bud and terminal bud in induced by EAC. Fijesh (2011), reported that MS medium supplemented with 2 mg/L BAP the extracted cells showed membrane blebbing, (Plate 1 and 2). Results revealed that nodal exvacuole formation and nuclear condensation plant with axillary buds possess comparatively which was absent in untreated cells. Thus, the higher survival chance. cytotoxic and antitumor effects of the leaf ex-

		LEAF		STEM	
Concentration	Standard	DLA	EAC	DLA	EAC
μg/ml	61.908	41.9	46.2	35.6	39.8
500 ^{µg/ml}	86.39	66.9	73.9	49.2	60.3
1000 µg/ml	98.19	78.2	86.4	62.1	79.4

Table 2. In vitro Anticancer Activity in Amaranthus viridis



Figure 1. Reducing sugar of Amaranthus viridis



Figure 2. Total carbohydrates and Total protein in Amaranthus viridis







Figure 4. Starch in Amaranthus viridis



Figure 5. Non-Enzymatic Antioxidants in Amaranthus viridis



Figure 6. Enzymatic Antioxidants in Amaranthus viridis



Figure 7. Invitro anticancer activity in Amaranthus viridis



Plate 1. inoculated explant of Amaranthus viridis



Plate 2. Node with axillary bud

Conclusion

diseases like asthma, gastrointestinal symptoms, for identifying the plant bio resource and its efand hepatic and cardiovascular disease. Present plants in culture can provide basis for future preliminary qualitative analysis of phytochemi- suitable explant of the plant for future studies. In cals, quantitative analysis of nutritional and anti- brief the wild species of Amaranthus viridis fits oxidant properties, anticancer analysis on DLA its claims of nutritional and medicinal properties and EAC cell lines and *in vitro* conservation, which satisfy its use as an ethnomedicinal plant Anticancer analysis on the crude methanolic ex- by tribal communities of Kerala. tract was carried out. Results obtained in the present study demonstrated the concentration References dependent anticancer effect of methanolic ex- Alegbejo J. O. 2013. Nutritional value and utilization of

cell lines which was found to be moderate to high. The results from the current study indicated that methanol extract of the Amaranthus viridiscontained various types of compounds with potential pharmacological activity. The presence of various bioactive compounds justifies the use of Amaranthus viridis for various ailments by traditional practitioners. From data, identification of more compounds in their extract and it previously reported that these compounds have antibacterial, antifungal, antioxidant and anticancer activity but further research should be made to isolate and purify natural products in their extract. In vitro conservation of the nodal explants of Amaranthus viridis was done with different explants (node with axillary bud and terminal bud) explants were inoculated in MS medium supplemented with BAP and the culture was maintained at 16 hours' photoperiod at a temperature of 26°C. This generated information on phytochemical, nutritional, and me-Tribal communities and local people of Kerala dicinal characteristics and therapeutic potential use the leaves of Amaranthus viridis for treating of Amaranthus viridis provide scientific proof skin disorders, respiratory and urinary problems fective utilisation in the future. The behaviour of study on Amaranthus viridis is concerned with conservation strategies and aid in selection of

tract of Amaranthus viridis in DLA and EAC Amaranthus (Amaranthus sps.)-a review. Bayero Journal of Pure and Applied Sciences, 6(1), 136-143.

ISSN 2583-0260 (online)

2013. Elemental, amino acid and phytochemical constitu- biochemistry 23. ents of fruits of three different species of eggplants. International Journal of Medicinal and Aromatic Plants; 3(2): Miller, G.L. 1972). Use of dinitrosalvcylic acid reagent for 200-202.

Bates L. S., Waldren R. A. and Teare I. D. 1973. Rapid determination of free proline for water-stress studies. Mishra, V. K., Bacheti, R. K. and Husen, A. (2011). Plant and soil, 39, 205-207.

Bendich, A. D. and Olson, J. A. (1989). Biological actions 77-196. of carotenoids 1. The FASEB journal. 3(8): 1927-1932.

David, Heber. and Lu, Q.Y. 2002). Overview of mechanisms of actionoflycopene Experimentalbiologyandmedicine. 227 (10):920-923.

Eom,S.H.,Park,H.J.,Jin,C.W.,Kim,D.O.,Seo,D.W.andJeo ng,Y.H.(2008). ChangesinantioxidantactivitywithtemperatureandtimeinChrysanthemumindicumL.teasduringel utionprocessesinhotwater. FoodScienceandBiotechnology .17:408-412.

Estebauer, H., Dieber, R. M., Striegl, G. and Waeg, G. 1991. Role of Vitamin-E inpreventing the oxidation of low density lipoprotein. American Journal of Clinical-Nutrition.53:314-321.

Gong, H., Zhu, X., Chen, K., Wang, S. and Zhang, C. (2005). Silicon alleviatesoxidativedamageofwheatplantsinpotsunder drought. PlantScience. 169: 313-321.

Hedge J.E., Hofreiter B.T. (1962). In: Methods in Carbohydrate Chemistry. (Eds. Whistler R L and Be Miller J N). Academic Press, New York; p. 17: 420.

Harborne J.B. (1977). Phenolic glycosides and their natural distribution in the biochemistry of phenolic compounds. Academic Press, New York, London; p. 152-16.

Lowry, O. H., Rosebrough, N. J., Farr. A. L.and Randall. R. J. (1951). Proteinmeasurementwithfolinphenolreagent.JournalofBiologicalChemistry.193:265-275.

Martin F. W., Telek L., Ruberte R. and Santiago A. G. 1979. Protein, oil and gossypol contents of a vegetable curd made from okra seeds. Journal of food Science, 44 (5), 1517-1519.

Martirosyan D. M., Miroshnichenko L. A., Kulakova S. N., Pogojeva A. V. and Zoloedov V. I. 2007. Amaranth oil application for coronary heart disease and hypertension. Lipids in health and disease, 6, 1-12.

Manthey, F. A. and Xu, Y. (2009). Glycobiology of Foods: Food Carbohydrates-Occurrence, Production,

Amadi B., Onuoha N., Amadi C., Ugbogu A., Duru M. Food Uses, and Healthful Properties. Advances in food

determination of reducing sugars. Analytical chemistrv.31:426-428.

Medicinal uses of chlorophyll: Acriticaloverview.Chlorophyll:Structure,functionandmedicinaluses:1

Pradeesh S. and Praveena P. (2020). Phytochemical investigation and in vitro anticancer activity of Sesbania grandiflora (L.) Pers. - A wild leafy vegetable of Southern Western Ghats. Journal of Advanced in Biological Science. 7(1&2): 49-53.

Pradeesh, S. and Swapna, T.S. (2018). Invitro studies and phytochemical evaluation of Bidensbiternata. LAPLAM-BERT, Academic Publishing:64-105.

Shingo, Y., Sakurada, S. and Nagumo, M. (1994). Role of intra cellular SOD in protecting human leukemic and cancer cells against superoxide and radiation. FreeRadicalBiologyandMedicine.17(5):389-395.

Szatmári S and Whitehouse P. Cochrane Dementia and Cognitive Improvement Group. 1996. Vinpocetine for cognitive impairment and dementia. Cochrane Database of Systematic Reviews, 2010(1).

Thayumanavan, B. and Sadasivam, S. (1984). Physicochemical basis for the preferential uses of cert ainricevarieties. QualityPlantFoodsforHumanNutrition.3 4:253.

Witham, F.H., Blaydes, D.F. and Devlin. R.M. (1971). Experiments in Plant Physiology, Van-Nostr and Reinhold Company, NewYork, USA.245.