# PHARMACOGNOSTIC PROFILING AND PHYTOCHEMICAL EVALUATION OF AZIMA **TETRACANTHA LAM. (SALVADORACEAE)**

# Supriya P. S.<sup>\*</sup> & Pillai Lakshmi Sreekumar

**Received: 19/4/2020** Accepted 25/5/2020

### Abstract

Standardization in medical practice is a system which ensure that every packet of medicine that is sold maintains the quality and quantity of the requisite bioactive compound and will induce the desired therapeutic effect. Azima is an important medicinal shrub used both in traditional system of medicine as well as in Ayurveda. The present investigation is concerned with the pharmacogonostic standardization and phytochemical screening of a very common species of Azima, Azima tetracantha. A morphological study of the plant was conducted. The whole plant was shade dried and ground to fine powdered and subjected organoleptic, fluorescent and physicochemical analysis. The plant powder was subjected to Soxhlet extraction using methanol. Phytochemical screening (Qualitative, Quantitative) of the methanol extract of the plant sample was conducted according to standard biochemical procedures. Antioxidant activity of the plant extract was tested using ABTS assay. The organoleptic analysis revealed the characteristic color, taste, odor and nature of the powder of Azima tetracantha. In fluorescence analysis, on treatment with different solvents, color changes could be noticed in the plant powder. The results of the physicochemical analysis provide an important parameter in detecting adulteration or improper handling of drugs. Qualitative and quantitative analysis confirmed the presence of many important phytochemicals in the plant. Bioactivity study indicated the anti-oxidant potential of the plant comparable to that of the standard Trolox. Further work should be carried out to isolate, purity and characterize the active constituents responsible for the specific activity of the plant. Also, additional work is necessary to elucidate the possible mechanism of action of the extract.

Key words: , Azima, Pharmacognosy, Fluorescence, Physio chemical, DPPH

## Introduction

traditional medicine, in the last few decades have 1914). been the subject for very intense pharmacological studies, this has been brought about by the acknowledgement of the values of medicinal plants as potential sources of lead compounds in drug development. However, mindless and unplanned felling of trees, shrubs and herbs has resulted in the loss of existing biodiversity (Gayathri et al., 2012).

tional medicine practices in the remote villages of Almost all the studies on A. tetracantha were

South Kerala. The plant is reported to have antispas-The importance of plants to humans and just about modic, diuretic, analgesic, wound- healing, antiall other life on Earth is fascinating. Plants fulfil the inflammatory, disinfectant, odontalgic, tonic, stimuthree basic needs of man such as food, clothing and lant, anti-diabetic, anti-diarrhoeal, anti- arthritic, exshelter. They are also sources of fuel, building mate- pectorant, anti-rheumatic, anti- catarrhal, antiperiodrials, craftwork material, dyes, transportation, rituals ic and astringent properties. It is used to get relief and medicine. Evidently, over the years, man has from muscular rheumatism and in diarrhoea and is been consistently exploiting plants for his various also given in dropsy, to relieve cough of phthisis and needs. Medicinal plants which form the backbone of asthma; for ulcers and in small pox (Rama Rao,

## **Relevance of the present study**

Traditional medicines still remain the main recourse for a large majority of people for treating health problems. Most of the plants used in herbal medicine practices, used by plant healers of remote villages and primitive aborigines have not yet been completely investigated for their phytochemical constituents The candidate plant, in the present investigation, Azi- and pharmacological activities. Azima tetracantha ma tetracantha Lam. is a member of the family Sal- Lam. is one such plant used in the folklore herbal vadoraceae (Mustard tree family). It is used in tradi- medicine practices in the villages of South Kerala.

Department of Botany, N.S.S. College Pandalam, Pathanamthitta, Kerala, India, 689501 *email* : Corresponding author<sup>\*</sup>: suprivasuresh634@gmail.com

conducted in the recent past (later than 2000). There- the fluorescence under normal light, short UV (100 fore, it appears that researchers became aware of the nm) and long UV (320 nm) was observed (Chase tremendous medicinal potential of A. tetracantha and Pratt, 1949). quite recently. Evaluation of the biopotential of A. tetracantha, and its phytoconstituents are scarcely Physicochemical characterization reported from plant materials growing in the state of Different physcio-chemical parameters were deter-Kerala. Azima tetracantha is very prevalent in the mined according to the official methods and guideadjacent state of Tamil Nadu and a few reports on lines on quality control for medicinal plant materials. the phytoconstituents and bioactivity have been reported from Tamil Nadu but not so from Kerala.

# **Materials and Methods**

## **Plant Material**

The plant selected for study, Azima tetracantha\_were collected from Chithara, Kollam. The specimen was authorized by Curator, Department of Botany, University of Kerala, Thiruvananthapuram.

## I. Morphological Study

The morphology of the species Azima tetracantha was studied using taxonomic kit based on qualitative and quantitative characters. The observations were recorded.

## **II.** Powder analysis

Fresh plant of Azima tetracantha\_is collected in polythene bag. The collected materials were washed under running tap water to remove adhered dirt. It was then rinsed with distilled water, blotted and B. Quantitative analysis (Harbone, 1973). dried in shade. The shade dried specimen was powdered in a mixer and sieved with fine mesh sieve. The powder was then used for the organoleptic study and solvent extraction.

## **Organoleptic study**

Organoleptic (literally '[impression on the organs') refers to evaluation by means of the organs of sense and includes the macroscopic appearance of the plant material, its color, odour and taste, occasionally the sound to 'snap' of its facture and the 'feel' of the powder to the touch (Wozniak et al., 1997). The plant powder characteristics like the colour, odour, taste and nature were evaluated.

## **Fluorescence analysis**

The crude drug powder was treated as such with eight different reagents solvents used were water, hydrochloric acid, sulphuric acid, sodium hydroxide, C. Quantitative analysis nitric acid, acetic acid, and methanol. Each solution was loaded on an activated thin gel layered slide and

- 1. Loss on drying (Indian Pharmacopoeia, 1992)
- 2. Foaming index (WHO, 1992)
- 3. Swelling index (WHO,1992)
- 4. Foreign matter (Indian Pharmacopoenia, 1996)
- 5. pH (Iqbal et al.,2010)

## **III. Phytochemical screening**

# A. Preparation and yield of extract (Indian Pharmacopoeia, 1996)

About 15 gm of the powdered plant material was subjected to extraction by Soxhlet apparatus using 100ml of methanol. The extract was concentrated under reduced pressure and preserved in refrigerator until further use. The percentage of the crude extract was determined using the following equation:

Percentage yield (%) = Weight of the crude extract x 100 weight of the sample

- 1. Tannins
- 2. Saponins
- 3. Flavonoids
- 4. Alkaloids
- 5. Terpenoids
- 6. Phlobatannins
- 7. Glycosides
- 8. Simple phenolics
- 9. Coumarins
- **10. Ouinones**
- 11. Acids
- **12. Flavanols**
- 13. Lignin
- 14. Steroids
- 15. Gums and mucilage

## 1. Determination of Alkaloids

2003)

**Determination of phenols** 3. (Spanos and Wrolstad, 1990

**Bio-activity study**—Anti-oxidant activity

### (3-ethylbenzthiazoline-6-sulphonic 1.Azino-bis acid) (ABTS) radical scavenging activity (Miller B. Quantitative analysis and Rice-Evans, 1997)

trations. The ABTS substrate working solution was compounds were present in the extract. Coumarins, prepared by adding 25µl of 3% hydrogen peroxide glycosides, quinones and mucilage were absent in solution to 10 ml of ABTS substrate solution. The the extract of the plant. The phytochemical screening assays were prepared in 96 wells plate. In the wells tests are provided (Table 4 and Plate 2.) for the Trolox standard, 10µl of Trolox and 20µl of Myoglobin working solution were added. In the C. Quantitative analysis wells for the test samples, 10µl (20-80µg/ml concen- The quantitative estimation of three phytochemical trations) of plant extracts and 20µl of myoglobin was done in the methanol extract of Azima tetracanworking standard were added. Then solution was tha (Table 5) by standard procedure. The amount of incubated for 5 minutes at room temperature. The phenols was the highest and that of alkaloids was stop solution (100µl) was added to each well. The low. end point absorbance was read at 405 nm using a plate reader

Percentage of inhibition (%) = Abs <sub>control</sub>-Abs <sub>sample</sub> x100 Abs control

## **Result and Discussion** I. Morphological study

An account of the morphological characters of Azima tetracantha Is given below (Table 1).

# **II.** Powder analysis

A. Organoleptic study Colour – Light olive green

Smell – Pungent smell Taste – Bitter Texture – Smooth

# **B.** Fluorescence analysis

The dry powder was subjected to fluorescence analysis with different reagents in normal light, short UV and long UV. The colour changes are summarised (Table2).

# C. Phytochemical characterization

A total of five physio-chemical parameters were evaluated in Azima tetracantha (Table 3). The mois-

ture content was reported to be in low amounts in the 2. Determination of steroids (Okeke and Elekwa, plant. Forming index was found to be more than 100 units and no considerable swelling was observed.

# I. Phytochemical screening

# A. Yield of the extract

The methanol extract was prepared by Soxhlet extraction. The yield of the methanol extract was 5.5%.

A total of 15 phytochemical were qualitatively ana-Trolox standards were prepared of different concen- lysed in methanol extract of the plant. Most of the

# D. Bioactivity study— Anti-oxidant activity

### (3-ethylbenzthiazoline-6-sulphonic 1.Azino-bis acid) (ABTS) radical scavenging activity (Miller and Rice-Evans, 1997)

The scavenging activity of the extract was compared with that of Trolox standard and percentage inhibition was calculated (Graph 1). The percentage inhibition of the ABTS radical by the methanol extract increased with increase in concentration. In this assay, by Probit analysis the methanol extract of Azima had good ABTS radical scavenging activity with  $IC_{50}$  value of 39.45µg/ml. the  $IC_{50}$  value of the standard Trolox was 36 µg/ml. The results are provided (Table 6 and Plate 3).

# Acknowledgement

The authors would like to thank Dr.G. Pramod, the principal, N.S.S. College Pandalam and Dr. G. Presanna Kumar, the head of the department, Post Graduate Department of Botany for helping to carry out this project.

Character	Description
Habit	Scrambling deciduous, shrub with axillary spines
Stem	Green herbaceous, branchlets branchlets terete or quadrangular. Young shoots pubescent, glabrous afterwards spines in each axial
Leaf	Greyish-green shining leaves with 2-3 pairs of prominent nerves from the base, spiny on tip. Leaves decussately opposite, simple and entire
Root	Taproot system.
Inflorescence	axillary, sometimes terminal spike or cyme up to 3cm long or flowers solitary; bracts ovate, often with long and spinous micro flowers.
Flower	unisexual, regular, tetramerous, usually sessile, small, axillary, greenish white.
Sepal	4 toothed calyx campanulate, 2–4mm long, with triangular lobes;
Petal	4, linear-oblong to oblong, greenish to yellowish, the upper part reflexed over the calyx
Androecium	male flowers with stamens inserted at the base of the rudimentary ovary, exerted; female flowers with staminodes
Gynoecium	superior ovary, up to 4.5mm long with a broad sessile stigma
Seed	Disc shaped, Black
Fruit	Globose, berry, greenish-white. Edible

Table 1. Morphological characters of         A	1zima tetracantha
--	-------------------

 Table 2. Fluorescent analysis of Azima tetracantha

Powder + Reagent	Visible (400- 800 nm)	U.V short (254 nm)	U.V long (366 nm)
Powder as such	Green	Black	Green
Powder + 1 N NaOH in $H_2O$	Black	Black	Dark green
Powder + 1N HCL	Black	Black	Dark green
Powder + 50% HNO <sub>3</sub>	Dark black	Black	Green
Powder + Iodine solution	Dark black	Black	Green
Powder + Acetic acid	Yellow	Black	Green
Powder + 5% FeCl <sub>3</sub>	Brown	Black	Bright green
Powder + Con. $H_2So_4$	Brown	Black	Bright green
Powder +Con. HCl	Brown	Black	Bright green
Powder + 95% Alcohol	Yellow	Black	Green

## Table 3. Physiochemical characters of Azima tetracantha

Parameters	Values
Foreign matter	2.4
Loss on drying	9%
Swelling index	3.79%
Foaming index	>100 Units
Ph	7.25

Table 4. Phytochemicals tested in Azima tetracantha

Sl.No	Phytochemicals	Present/Absent
1	Tannins	+
2	Saponins	+
3	Flavonoids	+
4	Alkaloids	+
5	Terpenoids	+
6	Phlobatannins	+
7	Glycosides	-
8	Simple phenolics	+
9	Coumarins	-
10	Quinones	-
11	Acids	+
12	Flavanols	+
13	Lignin	+
14	Steroids	+
15	Gums and mucilage	-

Table 5. Quantitative estimation of Azima tetracantha	Table 5. (	<b>Duantitative</b>	estimation	of Azima	tetracantha
---	------------	---------------------	------------	----------	-------------

Phytochemicals	Amount(mg/g)	
Alkaloids	0.40	
Steroids	10.4	
Phenols	85	

Table 6. ABTS	antioxidant assa	ιy
---------------	------------------	----

SL. No	Concentra- tion of extract	Trolox (percentage of inhibi- tion)	Plant extract (percentage of inhibition)
1	20µg/ml	41%	37%
2	40µg/ml	62%	59%
3	60µg/ml	88%	74%
4	80µg/ml	98%	96%

## References

Ameenah GF. 2006. Medicinal Plants-Tradition of yesterday and drugs of tomorrow, Molecular aspects of Medicine .27:1-93.

Ansari MM, Ahmad J, Ahmad A, Ansari SH. 2006. Pharmacognostic characterization and Standardization of *Morus alba* stem bark. *Journal of Medicinal and Aromatic Plant science* 28 (1): 31-36.

Antonisamy P, Duraipandiyan V and Ignacimuthu S. <u>2011</u>. Anti-inflammatory, analgesic and antipyretic effects of friedelin isolated from *Azima tetracantha* Lam. in mouse and rat models. *Journal of Pharmacy and Pharmacology* 63: 1070–1077.

Asiquith TN, Butter LG. 1986. Interaction of condensed tannins with selected proteins. *Phytochemistry* 25(7): 1591-1593.

Asolkar CV, Kakkar KK and Chakre OJ. 2000. Second supplement to Glossary of Indian Medicinal Plants with Active Principles Part I, National Institute of Science Communications, CSIR, New Delhi.

Begum NT, Ilyas MMH, Burkanudeen A, Kalavathy S, Vijayaanand A, Sampathkumar P and Jaswanth A. 2009. Hypoglycemic and antihyperlipidemic activity of ethanolic leaf extract of *Azima Tetracantha* Lam. on alloxan-induced diabetic rats. *Journal of Cell and Tissue Research* 9(1):1681-1685.

Chase CR and Pratt RJ. 1949. Fluorescence of powdered vegetable drugs with particular reference to development system of identification. *Journal of American Pharmacist Association* 38 (6): 324-331.

Chopra RN, Nayar SL and Chopra IC. 1956. Glossary of Indian Medicinal Plants, Council of Scientific and Industrial Research, New Delhi. p.32.

Cowan MM.1999. Plant products as antimicrobial agents. *Clinical Microbiology Reviews* 12(4): 564-582.

Cruz ASP. 2008. Anthelmintic effect of *Solanumlycocarpumin* mice infected with *Aspiculuristetraptera*. *The Journal of American Science* 4(3): 75-79

De Lucca A, Cleveland T, Rajasekhara K, Bouc S, Brown R. 2005. Fungal properties of CAY-1, a plant saponin for emerging fungal pathogens. 45<sup>th</sup> Interscience conference in Antimicrobial Agents and Chemotherapy Abstract 490: 180.